

**Linearising DNA: Before you start**

Measure the concentration of DNA in your final targeting construct DNA samples.

Prepare a 5µg sample of each final targeting construct.

Defrost BSA & NEB buffer 3. Place AsiSI enzyme on ice.

**Things you'll need**

AsiSI enzyme (NEB: R0630L, includes buffer and BSA)

10x Buffer 3 (NEB)

100x BSA (NEB)

37°C incubator

P20, P200 Gilson pipettes and filtered tips

MilliQ or Tissue Culture (TC) grade water

Ice

**Linearising DNA for electroporation**

- Decide on a final reaction volume (*Sanger uses 100µl*)
  - To 5µg of the final targeting construct add:
    - 1x Buffer 3 (*10µl*)
    - 1x BSA (*1µl*)
    - 4U/µg AsiSI enzyme (*20U = 2 µl of stock enzyme at 10,000U/ml*)
  - Add MilliQ or tissue culture (TC) grade water to the final required reaction volume
  - *Mix well*
  - Incubate overnight (or a minimum of 4 hours) at 37°C.
  - Store at -80°C.
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**DNA precipitation: Before you start**

- Defrost the linearised final targeting constructs
- Set the centrifuge to chill to 4°C
- Make up 70% ethanol using 100% stock and MilliQ/TC water (*chill on ice*)
- Chill an aliquot of 100% ethanol on ice

**Things you'll need**

Ethanol (99.7-100% v/v)

MilliQ or tissue culture grade water

Tissue culture grade PBS

Tissue culture hood

P20, P200 Gilson pipettes and filtered tips

4°C centrifuge

Ice

**DNA precipitation** (*use good sterile technique throughout*)

- Add 2-3 volumes of 100% ethanol to the linearised final targeting constructs (*e.g. 200-300µl ethanol to a 100µl digestion*)
- Seal the tube or plate carefully to prevent evaporation
- Incubate on ice for a minimum of 30 minutes
- Spin for 15 minutes, 3700rpm at 4°C
- Discard ethanol and check for evidence of precipitated DNA

*(white precipitate in bottom of well)*

- Add 200µl of 70% ethanol, spin plate for 5 minutes, 3700rpm, 4°C
- Discard ethanol
- Add 200µl of 70% ethanol, spin plate for 5 minutes, 3700rpm, 4°C
- Discard ethanol
- Add 200µl of 100% ethanol, spin plate for 5 minutes, 3700rpm, 4°C
- Discard ethanol
- Place samples in tissue culture hood and allow to dry for 5-10 minutes,
- Add 110µl PBS, seal vessel, label and chill overnight at 4 degrees
- Store linearised, precipitated DNA at -80°C until required for electroporation